
MOUSE/RAT EOSINOPHIL CATIONIC PROTEIN (ECP) ELISA KIT

FOR THE QUANTITATIVE DETERMINATION OF MOUSE / RAT EOSINOPHIL CATIONIC PROTEIN (ECP) CONCENTRATIONS IN SERUM AND EDTA PLASMA



FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

PURCHASE INFORMATION:

ELISA NAME	MOUSE/RAT EOSINOPHIL CATIONIC PROTEIN (ECP) ELISA
Catalog No.	SK00128-03
Lot No.	
Formulation	96 T
Standard range	156-10000 pg/ml
Sensitivity	50 pg/ml
Sample require	100 μl
Dilution Factor	Optimal dilutions should be determined by each laboratory for each application
Sample Type	Serum, EDTA Plasma
Specificity	Mouse , Rat
Intra-assay Precision	4-6%
Inter-assay Precision	8-12%
Storage	2-8°C

ORDER CONTACT:

AVISCERA BIOSCIENCE INC. 2348 Walsh Ave., Suite C Santa Clara, CA 95051 USA

Tel: (408) 982 0300 Fax: (408) 982 0301

Email: Info@AvisceraBioscience.com

www.AvisceraBioscience.com

INTRODUCTION

Mouse Eosinophil Cationic Protein (ECP) immunoassay is a 3.5 - 4.5 hour solid phase ELISA designed to measure Mouse Eosinophil Cationic Protein (ECP) in serum and plasma. It contains Mouse Eosinophil Cationic Protein (ECP) from Mouse Eosinophils and antibodies raised against recombinant Mouse ECP protein. Results obtained with naturally occurring Eosinophil Cationic Protein (ECP) samples showed linear curves that were parallel to the standard curves obtained using the kit standards. These results indicate that the immunoassay kit can be used to determine relative mass values for natural Mouse Eosinophil Cationic Protein (ECP). The data also indicated that rat EDTA plasma and serum samples cross-react with mouse Eosinophil Cationic Protein (ECP) ELISA kit.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for Eosinophil Cationic Protein (ECP) has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any Eosinophil Cationic Protein (ECP) present is bound by the immobilized antibody. After washing away any unbound substances, a biotinylated antibody specific for Eosinophil Cationic Protein (ECP) is added to the wells. Following a wash to remove any unbound antibody reagent, a Streptavidin-HRP conjugate is added to the wells. After washing away any unbound enzyme, a substrate solution is added to the wells and color develops in proportion to the amount of Eosinophil Cationic Protein (ECP) bound in the initial step. The color development is stopped and the intensity of the color is measured.

LIMITATIONS OF THE PROCEDURE

- _ FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- _ The kit should not be used beyond the expiration date on the kit label.
- _ Do not mix or substitute reagents with those from other lots or sources.
- _ It is important that the Dilution Buffer selected for the standard curve be consistent with the samples being assayed.
- _ If samples generate values higher than the highest standard, dilute the samples with the appropriate Dilution Buffer and repeat the assay.

_ Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

_ This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors have been tested in the immunoassay, the possibility of interference cannot be excluded.

MATERIALS PROVIDED

DESCRIPTION	CODE	QUANTITY
Eosinophil Cationic Protein (ECP) Microplate - 96 well polystyrene microplate (12 strips of 8 wells) coated with a purified antibody against Mouse Eosinophil Cationic Protein (ECP).	128-03-01	1 plate
Eosinophil Cationic Protein (ECP) Standard — 40 ng/vial of Mouse Eosinophil Cationic Protein (ECP) in a buffered protein base with preservatives; lyophilized.	128-03-02	1 vial
Detection Antibody— 105uL/vial, 100-fold concentrated of a purified antibody against mouse Eosinophil Cationic Protein (ECP) with preservatives; lyophilized.	128-03-03	1 vial
Positive Control – one vial of mouse Eosinophil Cationic Protein (ECP), lyophilized	128-03-04	1 vial
Streptavidin HRP Conjugate - 160 µl/vial, 75- fold concentrated solution of Streptavidin-HRP conjugate	SAHRP	1 vial
Dilution Buffer – 60 mL of buffered protein based solution with preservatives	DB08	1 bottle
HRP Diluent Solution - 12 mL of buffered protein based solution with preservatives	DB01	1 bottle
Wash Buffer - 50 ml of 10- fold concentrated buffered surfactant, with preservative.	WB01	1 bottle
TMB Substrate Solution - 11 ml of TMB substrate solution	TMB01	1 bottle

Stop Solution - 11 ml of 0.5M HCL	S-STOP	1 bottle
Plate Sealer	EAPS	1 piece
Plastic Pouch	P01	1 piece

STORAGE

Unopened Kit: Store at 2 - 8° C for up to 6 months. For longer storage, unopened Standard, Positive Control and Detection Antibody Concentrate should be stored at -20 or -70 °C. Do not use kit past expiration date.

Opened / Reconstituted Reagents: Reconstituted Standard and Detection Antibody Concentrate Solution SHOULD BE STORED at -20 °C or -70°C for up to one month. Streptavidin-HRP Conjugate 75-fold concentrated and other components may be stored at 2 - 8°C for up to 6 months.

Microplate Wells: Return unused wells to the plastic pouch with the desiccant pack, reseal along entire edge of zip-seal. Microplate may be stored for up to 6 months at 2 - 8° C.

OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 540 nm or 570 nm.
- Microplate shaker (250-300rpm).
- Pipettes and pipette tips.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinders.

PRECAUTIONS FOR USE

All reagents should be considered as potentially hazardous. The stop solution contains diluted hydrochloric acid. Appropriate care, therefore, should be taken while handling this solution. We therefore recommend that this product is handled only by those persons who have been trained in laboratory techniques and that it is used in accordance with the principles of good laboratory practice. Wear suitable protective clothing such as laboratory overalls, safety glasses and gloves. Care should be taken to avoid contact with skin or eyes. In the case of contact with skin or eyes wash immediately with water.

SAMPLE COLLECTION AND STORAGE

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at $1000 \times g$. Remove serum and assay immediately or aliquot and store samples at \leq -20° C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤-20° C. Avoid repeated freeze-thaw cycles.

Note: Use Aprotinin (enzyme inhibitor) (Code No.: 00700-01-25) for ALL sample collection to prevent sample degradation. 0.5 TIU per ml of sample solution.

SAMPLE PREPARATION

Mouse EDTA plasma or serum samples may require 4-8 fold dilution. A suggested 4-fold dilution is 60 μ L sample + 180 μ L Dilution Buffer (DB08). A suggested 8-fold dilution is 30 μ L sample + 210 μ L Dilution Buffer (DB08).

Rat EDTA plasma or serum samples may require 2 fold dilution. A suggested 2-fold dilution is 120 μ L sample + 120 μ L Dilution Buffer (DB08).

Optimal dilutions should be determined by each laboratory for each application. It is very important to pretest the sample dilution before performing the final assay.

Use polypropylene test tubes.

REAGENT PREPARATION

Bring all reagents to room temperature before use.

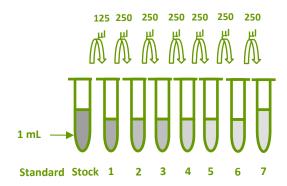
Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of Wash Buffer.

Eosinophil Cationic Protein (ECP) Standard - Refer to vial label for reconstitution volume. Reconstitute the Eosinophil Cationic Protein (ECP) Standard with 1.0 mL of Dilution Buffer. This reconstitution produces a stock solution of 40 ng/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 375 μ L of Dilution Buffer into tubes #1. Pipette 250 μ L of Dilution Buffer into tubes #2 to #7. Use the stock solution (40 ng/mL) to produce a dilution

series (below). Mix each tube thoroughly before the next transfer. The 10 ng/mL standard serves as the high standard. The Dilution Buffer serves as the zero

standard (0 ng/mL).

TUBE	STANDARD	DILUTION BUFFER	CONCENTRATION
stock	powder	1 ml	40 ng/ml
#1	125 μl of stock	375µl	10 ng/ml
# 2	250µl of 1	250µl	5 ng/ml
#3	250µl of 2	250µl	2.5 ng/ml
# 4	250µl of 3	250µl	1.25 ng/ml
# 5	of 4 لبا250	250µl	0.625 ng/ml
# 6	of 5 اµ250	250µl	0.3125 ng/ml
#7	250µl of 6	250µl	0.156 ng/ml



Concentration 40 10 5 2.5 1.25 0.6250.3120.156 ng/ml

Detection Antibody - Reconstitute the Detection Antibody concentrated with 105 μ L of **Dilution Buffer** (DB08) to produce a 100-fold concentrated stock solution. Transfer 105 μ L of 100-fold concentrated stock solution to 10.395 mL of **Dilution Buffer** (DB08) to prepare working solution.

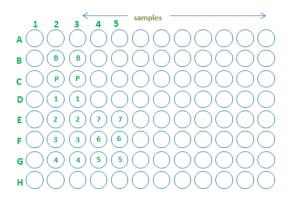
Streptavidin-HRP Conjugate - Transfer 160 μ l of 75-fold concentrated Streptavidin-HRP conjugate stock solution to 11.84 mL of HRP Diluent Solution (DB01) to prepare working solution. Note: 1x working solution of Streptavidin-HRP Conjugate should be used within a few days.

Positive Control- Reconstitute the Positive Control with 400 μ L of Dilution Buffer for 5-fold concentrated solution. Pipet 60 μ L of 5-fold concentrated solution into 240 μ L Dilution Buffer. Note: Positive Control should be prepared and used immediately.

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that blank, standards, positive control and samples be assayed in duplicates.

- 1. Prepare all reagents and working standards as directed in the previous sections.
- Remove excess micro-plate strips from the plate frame, return them to the plastic pouch with the desiccant pack, reseal.
- 3. Add 100 μ L of Dilution Buffer to Blank well (B2, B3).
- 4. Add 100 μL of Standard solution from #7 to #1 (reverse order of serial dilution) (E4, E5 to G4, G5 and G2, G3 to D2, D3), sample, or positive control per well (C2, C3). Cover with sealer. Incubate for 2 hours on micro-plate shaker at room temperature. A plate layout is provided to record standards and samples assayed.
- 5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Buffer (300 μL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
- 6. Add 100 μ L of Detection Antibody working solution to each well. Cover with sealer. Incubate for 2 hours on micro-plate shaker at room temperature.
- 7. Repeat the aspiration/wash as in step 5.
- Add 100 μL of Streptavidin-HRP Conjugate working solution to each well. Incubate for 60 minutes on micro-plate shaker at room temperature. Protect from light.
- 9. Repeat the aspiration/wash as in step 5.
- 10. Add 100 μ L of Substrate Solution to each well. Incubate for 6-9 minutes at room temperature. **Protect from light.**
- 11. Add 100 μ L of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green, or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
- 12. Determine the optical density of each well within 15 minutes, using a micro-plate reader set to 450 nm.



CALCULATION OF RESULTS

Average the duplicate readings for each standard, positive control, and sample and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a log-log curve fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the Eosinophil Cationic Protein (ECP) concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

TYPICAL DATA

This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

STANDARD (NG/ML)	AVERAGE OD450 (CORRECTED*)
Blank	0 (0.107)
0.156	0.042
0.313	0.092
0.625	0.176
1.25	0.341
2.5	0.667
5	1.208
10	2.224

CALIBRATION

This immunoassay is calibrated against a purified recombinant Mouse Eosinophil Cationic Protein (ECP).

SENSITIVITY

Twenty-five assays were evaluated and the minimum detectable dose (MDD) of Eosinophil Cationic Protein (ECP) was 50 pg/ml.

SPECIFICITY

This assay recognizes natural mouse Eosinophil Cationic Protein (ECP). The factors listed below were prepared at 1000 ng/mL in Dilution Buffer, and assayed for cross reactivity. The data also indicated that rat EDTA plasma and serum samples were bound to antibody that was used in this kit formulation condition. Its linear dilution curves were parallel to the standard curves obtained using the ELISA standard. That means rat EDTA plasma and serum samples cross-react with mouse Eosinophil Cationic Protein (ECP) ELISA kit.

PROTEIN NAME	CROSS-REACTIVITY (%)
Mouse Eosinophil Cationic Protein (ECP)	100
Mouse Periostin	0
Rat SPARC	0
Rat NGAL	0

SUMMARY OF ASSAY PROCEDURE

Add 100 µl of standard, samples, positive control to each well. Incubate 2 hours on plate shaker at RT. Aspirate and wash 4 times. Add 100 µl Detection Antibody Working Solution to each well. Incubate 2 hours on the plate shaker at RT. Aspirate and wash 4 times. Aspirate and wash 4 times. Add 100 µl Streptavidin-HRP conjugate Working Solution to each well. Incubate 60 min on the plate shaker at RT. Protect from light. Aspirate and wash 4 times. Add 100 µl Substrate Solution to each well. Incubate 6-9 min on the plate shaker. Protect from light. Add 100 µl Stop Solution to each well. Read 450nm

within 15 min