

HUMAN BONE MORPHOGENETIC PROTEIN 5 (BMP5) ELISA KIT

FOR THE QUANTITATIVE DETERMINATION
OF HUMAN BMP5 CONCENTRATIONS IN
CELL CULTURE SUPERNATES, EDTA PLASMA
AND SERUM



FOR RESEARCH USE ONLY. NOT FOR USE
IN DIAGNOSTIC PROCEDURES.

PURCHASE INFORMATION:

ELISA NAME	HUMAN BMP5 ELISA
Catalog No.	SK00013-01
Lot No.	
Formulation	96T
Standard range	93-6000 pg/mL
Sensitivity	30 pg/mL
Sample Volume	100 µl
Sample Type	Cell Culture Supernates, EDTA Plasma and Serum
Dilution Factor	(Optimal dilutions should be determined by each laboratory for each application)
Specificity	Human BMP5
Intra-assay Precision	4-6%
Inter-assay Precision	8-12%
Storage	2-8°C

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INTRODUCTION

Human BMP5 immunoassay is a 3.5 - 4.5 hour solid phase ELISA designed to measure human BMP5 in cell culture supernates, EDTA plasma and serum. It contains recombinant human BMP5 and antibodies raised against this protein. It has been shown to accurately quantify recombinant human BMP5. Results obtained with naturally occurring BMP5 samples showed linear curves that were parallel to the standard curves obtained using the kit standards. These results indicate that the immunoassay kit can be used to determine relative mass values for natural human BMP5.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for BMP5 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any BMP5 present is bound by the immobilized antibody. After washing away any unbound substances, a biotinylated antibody specific for BMP5 is added to the wells. Following a wash to remove any unbound antibody-biotin reagent, HRP link Streptavidin is added to the wells. After washing away any unbound enzyme, a substrate solution is added to the wells and color develops in proportion to the amount of BMP5 bound in the initial step. The color development is stopped and the intensity of the color is measured.

LIMITATIONS OF THE PROCEDURE

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_ The kit should not be used beyond the expiration date on the kit label.

_ Do not mix or substitute reagents with those from other lots or sources.

_ It is important that the Dilution Buffer selected for the standard curve be consistent with the samples being assayed.

_ If samples generate values higher than the highest standard, dilute the samples with Dilution Buffer and repeat the assay.

_ Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

_ This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors

have been tested in the immunoassay, the possibility of interference cannot be excluded.

MATERIALS PROVIDED

DESCRIPTION	CODE	QUANTITY
BMP5 Microplate - 96 well polystyrene microplate (12 strips of 8 wells) coated with a monoclonal antibody against BMP5.	013-01-01	1 plate
BMP5 Standard – 6000 pg/vial of recombinant human BMP5 in a buffered protein base with preservatives; lyophilized.	013-01-02	1 vial
Detection Antibody Concentrate – 110 µL/vial, 100-fold concentrate of biotinylated antibody against BMP5 with preservatives; lyophilized.	013-01-03	1 vial
Positive Control - one vial of recombinant human BMP5, lyophilized	013-01-04	1 vial
Streptavidin-HRP Conjugate - 75 ul/vial, 200-fold concentrated solution of Streptavidin conjugate to HRP	SAHRP	1 vial
Dilution Buffer - 60mL of buffered protein based solution with preservatives	DB01	1 bottle
Wash Buffer - 50 mL of 10-fold concentrated buffered surfactant, with preservative.	WB01	1 bottle
TMB Substrate Solution - 11 mL of TMB substrate solution	TMB01	1 bottle
Stop Solution - 11 ml of 0.5M HCl	S-STOP	1 bottle
Plate Sealer	EAPS	1 piece

STORAGE

Unopened Kit: Store at 2 - 8° C for up to 6 months. For longer storage, unopened Standard, Positive Control and Detection Antibody Concentrate should be stored at -20°C or -70°C. Do not use kit past expiration date.

Opened/Reconstituted Reagents: Reconstituted Standard and Detection Antibody SHOULD BE STORED at -20°C or -70°C for up to one month. Streptavidin-HRP Conjugate 200-fold concentrated

solution and other components may be stored at 2-8°C for up to 6 months.

Microplate Wells: Return unused wells to the plastic pouch containing the desiccant pack, reseal along entire edge of zip-seal. Microplate may be stored for up to 6 months at 2 - 8°C.

OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 540 nm or 570 nm.
- Microplate shaker (250-300rpm).
- Pipettes and pipette tips.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinders.

PRECAUTIONS FOR USE

All reagents should be considered as potentially hazardous. The stop solution contains diluted Hydrochloric acid. Appropriate care, therefore, should be taken while handling this solution. We therefore recommend that this product is handled only by those persons who have been trained in laboratory techniques and that it is used in accordance with the principles of good laboratory practice. Wear suitable protective clothing such as laboratory overalls, safety glasses and gloves. Care should be taken to avoid contact with skin or eyes. In the case of contact with skin or eyes wash immediately with water.

SAMPLE COLLECTION AND STORAGE

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

SAMPLE PREPARATION

Optimal dilutions should be determined by each laboratory for each application.

Use polypropylene test tubes.

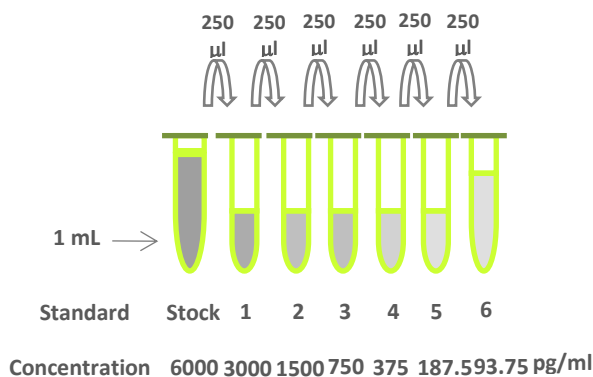
REAGENT PREPARATION

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of Wash Buffer.

BMP5 Standard - Refer to vial label for reconstitution volume. Reconstitute the **BMP5** Standard with 1.0 mL of Dilution Buffer. This reconstitution produces a stock solution of 6000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250 μL of Dilution Buffer into tubes #1 to #6. Use the high standard solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 6000 pg/mL standard serves as the high standard. The appropriate Dilution Buffer serves as the zero standard (0 pg/mL).

TUBE	STANDARD	DILUTION BUFFER	CONCENTRATION
stock	Powder	1000 μL	6000 pg/mL
# 1	100 μL of stock	400 μL	3000 pg/mL
# 2	250 μL of 1	250 μL	1500 pg/mL
# 3	250 μL of 2	250 μL	750 pg/mL
# 4	250 μL of 3	250 μL	375 pg/mL
# 5	250 μL of 4	250 μL	187.5 pg/mL
# 6	250 μL of 5	250 μL	93.75 pg/mL



Detection Antibody - Reconstitute the **Detection Antibody Concentrate** with 110 μ L of Dilution Buffer to produce a 100-fold concentrated stock solution. Pipette 10.89 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 110 μ L of 100-fold concentrated stock solution to prepare working solution.

Streptavidin-HRP Conjugate - Pipette 11.94 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 60 μ L of 200-fold concentrated stock solution to prepare working solution. *Note: 1x working solution of Streptavidin HRP Conjugate should be used within a few days.*

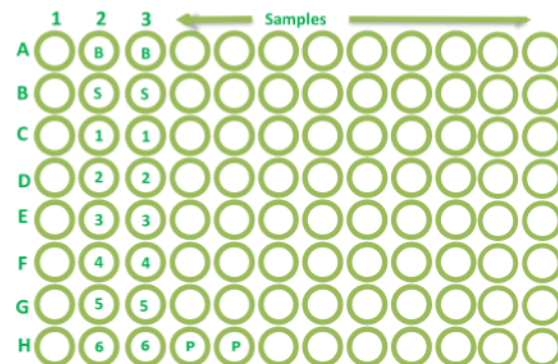
Positive Control - Reconstitute the **Positive Control** with 1.5 mL of Dilution Buffer. *Positive Control should be prepared and used immediately.*

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that standards be assayed in duplicate.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess micro-plate strips from the plate frame, return them to the plastic pouch containing the desiccant pack, reseal.
3. Add 100 μ L of Dilution Buffer to Blank well (A2, A3).
4. Add 100 μ L of Standard (B2, B3 to H2, H3), sample, or positive control (H4, H5) per well. Cover with sealer. Incubate for 2 hours on micro-plate shaker at room temperature. A plate layout is provided to record standards and samples assayed.
5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Buffer (300 μ L) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 100 μ L of Detection Antibody working solution to each well. Cover with sealer. Incubate for 2 hours on micro-plate shaker at room temperature.
7. Repeat the aspiration/wash as in step 5.

8. Add 100 μ L of **Streptavidin-HRP Conjugate** working solution to each well. Incubate for 60 minutes on micro-plate shaker at room temperature. **Protect from light.**
9. Repeat the aspiration/wash as in step 5.
10. Add 100 μ L of Substrate Solution to each well. Incubate for 12-18 minutes at room temperature. **Protect from light.**
11. Add 100 μ L of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green, or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
12. Determine the optical density of each well within 15 minutes, using a micro-plate reader set to 450 nm.



CALCULATION OF RESULTS

Average the duplicate readings for each standard, positive control, and sample and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a log-log curve fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the BMP5 concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

TYPICAL DATA

This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

BMP5 (PG/ML)	AVERAGE OD450 (CORRECTED)*
Blank	0 (0.091)
93.75	0.051
187.5	0.089
375	0.192
750	0.376
1500	0.631
3000	1.027
6000	1.692

*Lot No.:

** Positive Control: 100-200 pg/ml

CALIBRATION

This immunoassay is calibrated against a highly purified recombinant human BMP5.

SENSITIVITY

Twenty-five assays were evaluated and the minimum detectable dose (MDD) of BMP5 was 30 pg/mL.

SPECIFICITY

PROTEIN	CROSSREACTIVITY (%)
Human BMP5	100
Human BMP2	0
Human BMP4	0
Human BMP9	0
Human BMP7	0

SUMMARY OF ASSAY PROCEDURE**PREPARE REAGENTS, SAMPLES AND STANDARDS**

↓
Add 100µl of standard, samples, positive control to each well. Incubate 2 hours on the plate shaker at RT.

↓
Aspirate and wash 4 times.

↓
Add 100 µl Detection Antibody working solution to each well. Incubate 2 hours on the plate shaker at RT.

↓
Aspirate and wash 4 times.

↓
Add 100 µl Streptavidin-HRP conjugate working solution to each well. Incubate 60 minutes on the plate shaker at RT. **Protect from light.**

↓
Aspirate and wash 4 times.

↓
Add 100 µl Substrate Solution to each well. Incubate 12-18 min on the bench top. **Protect from light.**

↓
Add 100 µl Stop Solution to each well. Read 450nm within 15 min